

THE ROLE OF ROSE BENGAL TEST IN DETECTING HUMAN BRUCELLOSIS

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ABSTRACT

Brucellosis remains a serious public health concern, necessitating effective monitoring. This study evaluated the seroprevalence and identification of *Brucella* spp. in 300 blood specimens from patients, using phenotypic characterization and the Rose Bengal Test (RBT) for diagnosis. The percentage of positive serological tests was 9.3% (28/300), with a higher infection rate in females (10.0%) than in males (8.7%). The highest ratio was observed among patients aged 31-40 years (13.3%). Bacterial isolation confirmed *Brucella* species in 7.0% (21/300) of samples. Epidemiological analysis revealed a significantly higher prevalence among rural (10.6%) than urban (7.9%) participants and clear seasonality, with a peak incidence in September (12.0%). This study confirms the endemicity of brucellosis, establishes the RBT as a vital screening tool, and identifies key demographic and seasonal risk factors for guiding public health intervention strategies.

Keywords: Human Brucellosis, Prevalence, Risk Factors, RBT, Zoonosis.

INTRODUCTION

Human brucellosis is a bacterial infection that primarily affects humans and a variety of species, including domestic livestock, wildlife, rodents, and marine mammals. It is a highly prevalent zoonotic infection (Almashhadany *et al.*, 2026; WHO, 2020; Khairullah *et al.*, 2024; Heshteli *et al.*, 2025). The bacterium of the genus *Brucella* causes the condition. The bacteria are characterized as gram-negative intracellular coccobacilli, non-motile, and lacking spore formation. They are aerobic and urease-positive, and notably, they do not possess a capsule. Certain strains require a 5-10% CO₂ environment for optimal primary isolation (Hadad *et al.*, 1997; Bayhan *et al.*, 2020; Ötkün & Gürbilek, 2025; Shen *et al.*, 2025).

The infectious disease is human brucellosis, also known as Mediterranean fever, Malta fever, and undulant fever. Recently, the worldwide spread has undergone an important transformation due to a multifaceted interplay of factors, including animals, political instability, socioeconomic conditions, and inadequate sanitary controls (Almashhadany *et al.*, 2022; Bandara *et al.*, 2026; tkn & Grbilek, 2025). Brucellosis remains a persistent issue in many regions worldwide, including the Middle East, South and Central Asia, and Africa. This is mainly due to the lack of veterinary infrastructure and the widespread use of conventional livestock husbandry methods (Menshawy *et al.*, 2025). According to reports, Southern Europe has sporadic occurrences of the disease, whereas Northern and Western Europe have successfully eradicated it (European Health and Digital Executive Agency, 2022; Ram, 2025).

Several factors affect the prevalence of undulant fever, including susceptible animal hosts, social and economic conditions, cultural practices, and the implementation of public health interventions, as well as a combination of biological, environmental, social, and economic factors (Corbel, 2006; Dadar & Maurin, 2025). Cattle, sheep, goats, and swine serve as critical reservoirs, excreting *Brucella* spp. through milk, urine, excrement, and reproductive materials (Parsaeimehr *et al.*, 2026; Zeb *et al.*, 2025). The maintenance and dissemination of the infection to domestic populations depend on wildlife reservoirs, including elk, bison, and wild boar (Rebollada-Merino *et al.*, 2024; Wang *et al.*, 2025).

Consumption of unprocessed milk and milk products significantly contributes to the incidence of human brucellosis. Implementing effective control measures such as vaccination and strict food safety protocols is vital for reducing the burden of the illness (Mascolo *et al.*, 2026; Kithuka *et al.*, 2025). Official data from the World Health Organization (WHO) indicate an annual incidence of approximately 500,000 cases, yet it is widely regarded as significantly underestimating the true extent of infection (Mohammed & Girma, 2026; Fernandez-Georges *et al.*, 2025).

Humans can become infected through various pathways: by ingesting unprocessed dairy and milk derivatives; by inhaling contaminated aerosols in the environment, and via direct contact with infected animals and their products. The bacteria infiltrate via mucous membranes, damage skin lesions, proliferate in the lymph nodes, and cause bacteremia, thereby disseminating throughout the body. While infrequent, instances of transmission between individuals have been documented through breastfeeding, blood transfusions, bone marrow transplants, sharing of contaminated needles, and transplacental infection (Al-Abd *et al.*, 2025; Mazzeo *et al.*, 2025).

Clinical symptoms appear 2 to 4 weeks following infection, with an onset that may be gradual and insidious or notably acute. The symptom profile features intermittent fever, significant malaise, headache, myalgia, arthralgia, and respiratory or gastrointestinal issues such as cough and abdominal pain. Commonly observed are constitutional symptoms like anorexia, weight loss, and night sweats (Jin *et al.*, 2023; Ibrahim *et al.*, 2025; Hussein *et al.*, 2025).

The genus *Brucella* includes 12 known species. The first category includes six traditional species: *Brucella abortus*, *Brucella melitensis*, *Brucella suis*, *Brucella canis*, *Brucella ovis*, *Brucella neotomae*, and the second category includes six recently identified species: *Brucella ceti*, *Brucella pinnipedialis*, *Brucella microti*, *Brucella inopinata*, *Brucella papionis*, and *Brucella vulpis*. Each species exhibits a distinct preference for its host reservoir: cattle are the host for *B. abortus*, goats and sheep are considered hosts for *B. melitensis*, and *Brucella suis* is known to infect pigs. The significance of these three species lies in their global endemicity (Ferrero *et al.*, 2020; Rossetti *et al.*, 2022; Qureshi *et al.*, 2024; Han *et al.*, 2025).

Present diagnostic approaches integrate clinical assessment, which is nonspecific, with serological testing. Serological assays remain fundamental in routine diagnosis due to their affordability, ease of operation, and high sensitivity (Almashhadany *et al.*, 2023; Pang *et al.*, 2025). The serology test (RBT) is a quick test that depends **on** agglutination on a slide **using a colored** *Brucella abortus* antigen at an **acidic** pH of 3.6-3.7. Initially developed for veterinary screening, its straightforward nature, affordability, rapid results, and high precision have led to its widespread use as a preliminary test for identifying brucellosis in humans (Elbehiry *et al.*, 2023; Akhtardanesh *et al.*, 2025; Almuzaini *et al.*, 2025). Consequently, this study aims to: assess the prevalence of human brucellosis in Erbil Governorate, categorized based on habitation area (urban or rural), sex, age groups, and to evaluate the effect of seasonal conditions throughout the study period on the spread of this disease

METHODS

Collection of Specimens and Serological Identification

A total of 300 human blood specimens were randomly obtained. Data on participants registered by site of habitation, age, and sex were collected in Erbil City. The study was conducted from 1/7/2024 to 30/12/2024. 150 samples were obtained from each sex, with ages ranging from 1 to over 70 years. The samples included 140 blood specimens collected from an urban area and 160 from a rural area. The blood samples of each individual were collected in ten (10) ml increments. Each sample was divided into two tubes: Five milliliters were collected in a vacutainer tube that did not include any anticoagulants, and five milliliters were collected in a vacutainer tube that did contain anticoagulants. The collected specimens were transported in a sterile container to the Medical Microbiology Laboratory, Knowledge University/College of Science. The initial blood fraction was allowed to coagulate before centrifugation, which separated serum for Brucella antibody detection. The Brucella species were isolated using the second portion of the blood. (Al-mashhadany 2014).

Detection of Brucella Antibodies

The diagnosis of specific antibodies in patients was identified depending on the RBT TEST. According to Al Mashhadany (2018), mix 0.03 ml of test blood with an equivalent amount of antigen on an enamel plate, gently agitate for 4 minutes, and record agglutination as any perceptible reaction.

Isolation of Brucella Species

For Brucella species isolation, 5–7 mL of blood was inoculated into an aerobic blood culture bottle. Blood culture was performed using the BACTEC 9240 system (Becton Dickinson, Franklin Lakes, USA), and all bottles were incubated for up to 4 weeks. When a positive signal was detected, subculture onto Brucella base blood agar was performed; colonies were identified based on colony morphology and biochemical tests (oxidase, urease, catalase, nitrate reduction), as described elsewhere (Almashhadany et al., 2022). Cultures that did not produce a positive signal by the end of the fourth week were considered negative for Brucella spp.

Identification of Brucella Species

All *Brucella* spp. isolates were identified to the species level based on H₂S production, thionine sensitivity, CO₂ requirement, and agglutination with monospecific sera A and M (Hadad et al., 1997; Almashhadany et al., 2022).

Statistical Analysis

Data were analyzed using the chi-square test in SPSS version 15 to assess differences.

RESULTS

Detection of RBT Results

The results in Table 1 showed that 28 (9.3%) brucellosis cases were recorded among the 300 blood specimens assessed. The percentage of infections among female patients was 15 (10.0%), compared with 13 (8.7%) among male patients.

Table 1: Distribution of RBT among Human Samples According to Sex

Sex	No of Samples examined	Positive N (%)		Negative N (%)		X ²	P-Value
Male	150	13	8.7	137	91.3	0.158	0.691
Female	150	15	10.0	135	90.0		
Total	300	28	9.3	272	90.7		

RBT results among Human Samples based on Age

The high prevalence of brucellosis infection was recorded in two age groups, between (31- 40) and (21–30) years, with percentages of (13.3 %) and (12.5%), respectively, as shown in Table 2.

Table 2: Results of RBT Among Human Samples According to Age

Age Group (Years)	No. Examined	Positive N (%)		Negative N (%)		X ²	P-Value
1-10	30	2	6.7	28	93.3	2.572	0.922
11-20	35	3	8.6	32	91.4		
21-30	40	5	12.5	35	87.5		
31-40	45	6	13.3	39	86.7		
41- 50	35	4	11.4	31	88.6		
51 - 60	45	3	6.7	42	93.3		
61 - 70	40	3	7.5	37	92.5		
< 71	30	2	6.7	28	93.3		
Total	300	28	9.3	272	90.7		

Habitat distribution of Human Brucellosis

The data from our study revealed that the prevalence of brucellosis infection, based on habitation, was higher in rural areas than in urban areas, at 17/160 (10.6%) and 11/140 (7.9%), respectively (Table 3).

Table 3: Seroprevalence of Human Brucellosis in Human Blood Samples According to Residency

Source of samples	No examined	Positive N (%)		Negative N(%)		X ²	P-Value
Urban area	140	11	7.9	129	92.1	0.676	0.411
Rural area	160	17	10.6	143	89.4		
Total	300	28	9.3	272	90.7		

Brucella spp Identification

The results in Table 4 showed that, according to the morphological characteristics of growth on culture media and biochemical tests, *Brucella* species were isolated from 21 (7.0%) of 300 specimens. This study revealed that *Brucella abortus* was isolated from male 6/9 (66.7%) and female 7/12 (58.3%), while *Brucella melitensis* was isolated from male and female in ratios of 3/9 (33.3%) and 5/12 (41.7%), respectively.

Table 4: Isolation rate of Brucella species from Human Blood Samples

Sex	No. sample	Positive N (%)		Negative N (%)		<i>Br. melitensis</i> N (%)	X ²	P-Value
Male	150	9	6.0	6	66.7	3 33.3	0.205	0.651
Female	150	12	8.0	7	58.3	5 41.7		
Total	300	21	7.0	13	61.9	8 38.1		

Relation between seasonal conditions and seroprevalence.

The results, as shown in Table 5, illustrated differences in the seroprevalence of specific Brucella antibodies in patients' blood. High value of infection found in September 6/50 (12.0%), then in August 5/50 (10.0%), whereas the infection percentage reduction in December was 3/50(6.0%) and in July 4/50 (8.0%)

Table 5: Seasonal Variation of Brucellosis Seroprevalence (RBT)

Months	No. examined	RBT N (%)	X ²	P-Value
July	50	4 8.0	1.261	0.939
August	50	5 10.0		
September	50	6 12.0		
October	50	5 10.0		
November	50	5 10.0		
December	50	3 6.0		
Total	300	28 9.3		

DISCUSSION

Brucellosis illness is considered a persistent and important problem for human health that severely impacts global socio-economic progress, particularly in the developing world, as consistently reported by major international organizations (Franc *et al.*, 2018; Kanu, 2024), including the Food and Agriculture Organization (FAO) and the World Organization for Animal Health (OIE). World Health Organization (WHO), The serological test RBT is a widely adopted global tool for epidemiological studies that uses human sera due to its affordability, simplicity, sensitivity, and ease of use, making it exceptionally effective for the detection of specific bacterial antibodies (Di Bonaventura *et al.*, 2021; Islam *et al.*, 2025). The 9.3% value recorded in our research is consistent with Din *et al.* (2013), who reported a reach of 9.33% according to RBT. In comparison, Salem *et al.* (2014) in Egypt reported a prevalence of 11.1%, while Shome *et al.* (2017) in India reported a prevalence of 6.38%. In Angola, Mufinda *et al.* (2017) mentioned a general weighted percentage of 15.6%. The study found that females were more exposed (10.0%) than males (8.7%). This was compatible with Diju (2009) in Pakistan (25.58% in females vs 10.18% in males). Conversely, Salem *et al.* (2014) found lower rates in females (8.9%) than in males (11.6%), and Tumwine *et al.* (2015) also reported higher rates in males (20.5% vs 15.3%). However, our study revealed no statistically significant difference between sexes according to the chi-square test (0.158).

Regarding Table 2, the highest percentage was in patients aged 31–40 years, followed by 21–30 and 41–50 years (13.3%, 12.5%, and 11.4%, respectively), with no statistically significant variation in age-based risk according to the chi-square test ($\chi^2 = 2.572$). This aligns with Abdul-Razag (2015) in Yemen, who noted a high prevalence (17.4%) in the same age group among males. However, Riabi *et al.* (2017) in Iran reported a higher incidence in patients over 50 years old (30%).

In an investigation of brucellosis seroprevalence among Indian veterinary healthcare workers, Shome *et al.* (2017) analyzed 1,050 serum samples and found a significant gender-based risk. While males constituted the vast majority of the study group (94.5%), all seropositive cases were confined to this demographic, yielding a 7.46% infection rate and a statistically significant result ($p=0.029$). The current results were consistent with Sharma *et al.* (2016), who reported an overall seroprevalence of 4.96% and test-wise seroprevalences of 9.91% by RBPT, 9.91% by mRBPT, 9.09% by STAT, and 16.52% by I-ELISA. They reported that the prevalence of brucellosis was higher in the 35–50 years age group than in the 20–35 and 50–65 years age groups, and also confirmed that the sex-wise seroprevalence was higher in males than in females. This finding contrasts with research conducted in Uganda by Tumwine *et al.* (2015), which reported a higher seroprevalence of human brucellosis in individuals over 60 years old, at 22.2%. Additionally, the percentage among all patients in the younger age groups, specifically those with ages less than thirteen years, 13 to 19 years, and 20 to 59 years, was recorded at 16.7%. Furthermore, a recent study by Riabi *et al.* (2017) in Iran indicated that the majority of brucellosis cases occurred in patients older than 50 years (30%), while the lowest incidence 12.3 % was observed in individuals aged 31–40 years.

Nielsen and Gall (2001) indicated that the specificity and sensitivity of the RBT are reported to fluctuate with the serum hydrogen ion concentration (pH) and the surrounding temperature of the antigen. The principle of RBT operation depends on detecting immunoglobulins (Ig types: A, M, G1, G2) in the test serum and on a reduction in IgM activity at a lower pH value.

Based on the participants' living conditions (Table 3), a notable seroprevalence of *Brucella* infection was observed: 17 out of 160 (10.6%) in rural areas and 11 out of 140 (7.9%) in urban areas. Ebrahimpour *et al.* (2012) reported in Iran that out of 337 participants diagnosed with brucellosis, 25.72% were urban dwellers and 74.28% lived in rural settings. In a study by Dieckhaus and Kyebambe (2017) conducted in Uganda, it was noted that rural patients are often involved in livestock farming; additionally, drinking unpasteurized milk may be a major route of transmission. No variation in the percentage of humans was found by habitation (0.676).

Based on the research data (Table 4), the total isolation rate was 21 (7.0%) out of 300 specimens, depending on the morphological characteristics of *Brucella*. This finding revealed that the isolation rate in males and females for *Brucella abortus* was 6/9 (66.7%) and 7/12 (58.3%), while for *Brucella melitensis* it was 3/9 (33.3%) and 5/12 (41.7%), which showed no statistically significant variation among the isolation rates of *Brucella* species from human blood samples. Our findings were consistent with those of Minas *et al.* (2007) and Arshi *et al.* (2017), which reported isolation rates of 8.49% and 6.0%, respectively, for *Brucella* spp. At the same time, Ogola *et al.* (2014) reported a higher percentage than our findings in their study assessing the prevalence of disease in animals and humans living in the same house. They reported human infection rates ranging from 2.2% to 14.1% and animal infection rates ranging from 1.2% to 3.4%.

Another aspect of this study addresses the correlation between months and the prevalence of Brucella antibodies in human samples. The results presented in Table 5 indicate that the peak antibodies occurred in September (6 out of 50; 12.0%), followed by August (5 out of 50; 10.0%), while the lowest rates were recorded in December (3 out of 50; 6.0%) and July (4 out of 50; 8.0%), respectively, and no variation was found in the ratio of different months. However, bacterial transmission may be associated with contaminated food products such as meat and milk. Minas et al. (2007) reported that human brucellosis exhibits a strong seasonal pattern, with the majority of cases diagnosed between December and May. Riabi et al. (2017) in Iran found that, among 176 patients, 94.8% resided in rural areas and 5.2% in urban areas. The highest incidence of reported cases occurred in June and July, with the lowest figures recorded in January. It appears that the disease prevalence begins in weather with optimal temperatures (in spring), peaks in high-temperature environments (in summer), and shows a low disease prevalence in autumn.

CONCLUSION

Brucellosis remains a pressing public health challenge in Iraqi Kurdistan, necessitating a coordinated, multi-sectoral strategy. The Rose Bengal Test (RBT) is an ideal surveillance tool for human cases due to its affordability, simplicity, and high sensitivity. However, effective control extends beyond diagnostics; it requires sustained public awareness campaigns across all media platforms to educate communities about transmission pathways. Special emphasis should be placed on rural populations, where promoting the consumption of pasteurized milk and dairy products is essential. Ultimately, the long-term control of this zoonotic disease hinges on eliminating brucellosis in livestock, an indispensable foundation for safeguarding human health. A unified effort involving public health authorities, veterinary services, and community engagement will be key to reducing the burden of brucellosis in the region.

Conflicts of Interest

The authors declare no conflicts of interest.

REFERENCES

- Abdul-Razag, W. M. A. A. (2015). *Serological and bacteriological study on brucellosis in human and food-producing animals in Thamar Province* [Master's thesis, Dhamar University].
- Akhtardanesh, B., et al. (2025). Molecular and serological investigation of *Brucella* species in kennel and farm dogs in Iran. *Acta Tropica*, 262, Article 107521. <https://doi.org/10.1016/j.actatropica.2025.107521>
- Al-Abd, N., et al. (2025). Brucellosis: A comprehensive review of epidemiology, pathogenesis, diagnosis, treatment, and global prevalence. *Electronic Journal of University of Aden for Basic and Applied Sciences*, 6(2), 131–140. <https://doi.org/10.47372/ejua-ba.2025.2.448>
- Al-mashhadany, D. A. (2014). Prevalence of brucellosis in humans and camels in Thamar Province/Yemen. *Journal of the Saudi Society of Agricultural Sciences*, 13(2), 132–137.
- Al-mashhadany, D. A. (2018). Application of Rose Bengal Test for surveillance of human brucellosis in Erbil Governorate, Kurdistan Region, Iraq. *RJLBPCS*, 3(4), 162. <https://doi.org/10.26479/2018.0401.14>
- Almashhadany, D. A., et al. (2022). Epidemiological study of human brucellosis among febrile patients in Erbil-Kurdistan region, Iraq. *Journal of Infection in Developing Countries*, 16(7), 1185–1190. <https://doi.org/10.3855/jidc.15669>
- Almashhadany, D. A., et al. (2023). Reliable and highly specific techniques for the detection of *Brucella* spp. antibodies in camel milk. *Iraqi Journal of Veterinary Sciences*, 37(4), 795–800.
- Almashhadany, D. A., et al. (2026). Assessing human brucellosis infection rates in high-risk occupational groups. *Journal of Infection in Developing Countries*, 20, 79–86. <https://doi.org/10.3855/jidc.21171>
- Almuzaini, A. M., et al. (2025). Seroprevalence of brucellosis in camels and humans in the Al-Qassim region of Saudi Arabia and its implications for public health. *AMB Express*, 15(1), 22. <https://doi.org/10.1186/s13568-025-01822-8>
- Arshi, A., et al. (2017). Molecular method for direct detection of *Brucella* spp. in human blood samples. *Annual Research & Review in Biology*, 20(4), 1–7. <https://doi.org/10.9734/ARRB/2017/37771>
- Bandara, D. M. C. L., et al. (2026). Seroprevalence and risk factors for bovine brucellosis in Polonnaruwa District, Sri Lanka. *The Journal of Agricultural Sciences - Sri Lanka*, 21(1), 161–175. <https://doi.org/10.4038/jas.v21i1.11719>
- Bayhan, G. I., et al. (2020). Pulmonary infections due to brucellosis in childhood. *Tuberk Toraks*, 68(1), 43–47. <https://doi.org/10.5578/tt.69015>
- Corbel, M. J. (2006). *Brucellosis in humans and animals*. World Health Organization.
- Dadar, M., & Maurin, M. (2025). Commentary on: Brucellosis in Iraq: A comprehensive overview of public health and agricultural challenges. *German Journal of Microbiology*, 5(1), 1–3.
- Di Bonaventura, G., et al. (2021). Microbiological laboratory diagnosis of human brucellosis: An overview. *Pathogens*, 10(12), 1623. <https://doi.org/10.3390/pathogens10121623>
- Dieckhaus, K. D., & Kyebambe, P. S. (2017). Human brucellosis in rural Uganda: Clinical manifestations, diagnosis, and comorbidities at Kabale Regional Referral Hospital, Kabale, Uganda. *Open Forum Infectious Diseases*, 4(4), ofx237. <https://doi.org/10.1093/ofid/ofx237>
- Diju, I. U. (2009). Brucellosis--an under-estimated cause of arthralgia & muscular pains in the general population. *Journal of Ayub Medical College Abbottabad*, 21(2), 128–131.
- Din, A. M. U., et al. (2013). A study on the seroprevalence of brucellosis in human and goat populations of the district Bhimber, Azad Jammu and Kashmir. *The Journal of Animal and Plant Sciences*, 23(1 Suppl.), 113–118.
- Ebrahimpour, S., et al. (2012). The prevalence of human brucellosis in Mazandaran Province, Iran. *African Journal of Microbiology Research*, 6(19), 4090–4094. <https://doi.org/10.5897/AJMR11.1076>
- Elbehiry, A., et al. (2023). The development of diagnostic and vaccine strategies for early detection and control of human brucellosis, particularly in endemic areas. *Vaccines*, 11(3), 654. <https://doi.org/10.3390/vaccines11030654>

- European Health and Digital Executive Agency. (2022). *Brucellosis*. https://hadea.ec.europa.eu/programmes/single-market-programme-food/veterinaryprogrammes/brucellosis_en
- Fernández-Georges, I. K., et al. (2025). Ruminant and human brucellosis situation in Türkiye and the Caucasus. *Tropical Animal Health and Production*, 57, 296. <https://doi.org/10.1007/s11250-025-04525-1>
- Ferrero, M. C., et al. (2020). Pathogenesis and immune response in Brucella infection acquired by the respiratory route. *Microbes and Infection*, 22(9), 407–415. <https://doi.org/10.1016/J.MICINF.2020.06.001>
- Franc, K. A., et al. (2018). Brucellosis remains a neglected disease in the developing world: A call for interdisciplinary action. *BMC Public Health*, 18, 125. <https://doi.org/10.1186/s12889-017-5016-y>
- Hadad, J., et al. (1997). Isolation of Brucella strains from dairy products in Ninevah Province, Iraq. *Iraqi Journal of Veterinary Sciences*, 10(1), 39–44.
- Han, C., et al. (2025). Machine-learning-based pangenome-wide association studies reveal the impact of host source on the zoonotic potential of closely related bacterial pathogens. *Communications Biology*, 8, 1253. <https://doi.org/10.1038/s42003-025-08650-3>
- Heshteli, R. R., et al. (2025). Advances in biosensors: A breakthrough in rapid and precise brucellosis detection. *Analytical Biochemistry*, 700, Article 115782. <https://doi.org/10.1016/j.ab.2025.115782>
- Hussein, M., et al. (2025). Seroprevalence and molecular investigation of camel brucellosis in Egypt with emphasis on potential risk factors. *German Journal of Veterinary Research*, 5(2), 15–23. <https://doi.org/10.51585/givr.2025.2.0130>
- Ibrahim, I. I., et al. (2025). Uncommon presentation of neurobrucellosis. *Neurosciences (Riyadh)*, 30(3), 241–246. <https://doi.org/10.17712/nsj.2025.3.20240081>
- Islam, S., et al. (2025). Beyond serology: A meta-analysis of advancements in molecular detection of Brucella spp. in seronegative animals and biological samples. *Veterinary Medicine and Science*, 11, e70200. <https://doi.org/10.1002/yms3.70200>
- Jin, M., et al. (2023). Research progress on complications of brucellosis. *Frontiers in Cellular and Infection Microbiology*, 13, Article 1136674. <https://doi.org/10.3389/fcimb.2023.1136674>
- Kanu, S. (2024). The socio-economic impact of brucellosis outbreaks among large and small ruminants under an extensive nomadic management system in Sierra Leone. *IntechOpen*. <https://doi.org/10.5772/intechopen.114278>
- Khairullah, A. R., et al. (2024). Brucellosis: Unveiling the complexities of a pervasive zoonotic disease and its global impacts. *Open Veterinary Journal*, 14(5), 1081. <https://doi.org/10.5455/OVJ.2024.V14.I5.1>
- Kithuka, J. M., et al. (2025). The burden of brucellosis in donkeys and its implications for public health and animal welfare: A systematic review and meta-analysis. *Veterinary World*, 18(2), 368–379. <https://doi.org/10.14202/vetworld.2025.367-378>
- Lai, S., et al. (2017). Changing epidemiology of human brucellosis, China, 1955–2014. *Emerging Infectious Diseases*, 23(2), 184–194.
- Mascolo, C., et al. (2026). Evaluation of brucellosis eradication strategies in water buffalo in a key dairy production area of southern Italy. *Frontiers in Microbiology*, 17, Article 1741007. <https://doi.org/10.3389/fmicb.2026.1741007>
- Mazzeo, A., et al. (2025). Brucellosis in cattle and buffalo in southern Italian provinces: Trends in the presence of territory-specific One Health measures. *Frontiers in Microbiology*, 16, Article 1609336. <https://doi.org/10.3389/fmicb.2025.1609336>
- Menshawy, A. M. S., et al. (2025). Animal brucellosis in Egypt: Review on evolution, epidemiological situation, prevalent Brucella strains, genetic diversity, and assessment of implemented national control measures. *Microorganisms*, 13(1), 170. <https://doi.org/10.3390/MICROORGANISMS13010170>
- Minas, M., et al. (2007). Epidemiological and clinical aspects of human brucellosis in Central Greece. *Japanese Journal of Infectious Diseases*, 60(6), 362–366.
- Mohammed, H., & Girma, T. (2026). Assessment of knowledge, practice, and factors associated with brucellosis prevention among residents in Bati District, Oromiya Zone, Amhara State, Ethiopia. *New York Science Journal*, 19(1), 85–91. <https://doi.org/10.7537/marsnys19>

- Mufinda, F. C., et al. (2017). Prevalence and factors associated with human brucellosis in livestock professionals. *Revista de Saude Publica*, 51, 57. <https://doi.org/10.1590/S1518-8787.2017051006051>
- Nielsen, K., & Gall, D. (2001). Fluorescence polarization assay for the diagnosis of brucellosis: A review. *Journal of Immunoassay and Immunochemistry*, 22(3), 183–201. <https://doi.org/10.1081/IAS-100104705>
- Ogola, E., et al. (2014). Sero-prevalence of brucellosis in humans and their animals: A linked cross-sectional study in two selected counties in Kenya. *Online Journal of Public Health Informatics*, 6(1), e67. <https://doi.org/10.5210/ojphi.v6i1.5166>
- Ötkün, S., & Gürbilek, S. E. (2025). Brucella and brucellosis: Ancient origins, modern challenges. *Egyptian Journal of Veterinary Science*, 1–6. <https://doi.org/10.21608/ejvs.2025.366582.2685>
- Pang, C., et al. (2025). Delayed diagnosis of pediatric brucellosis: A case study and literature review. *Annals of Medicine and Surgery*, 87(6), 3880–3884. <https://doi.org/10.1097/MS9.0000000000003278>
- Parsaeimehr, M., et al. (2026). Detection of Brucella spp. in raw milk and dairy products of traditional domestic dairy sale centers by Real-Time PCR in Semnan. *Archives of Razi Institute*. <https://doi.org/10.22092/ari.2025.368595.3543>
- Qureshi, K. A., et al. (2024). Brucellosis: Epidemiology, pathogenesis, diagnosis and treatment—a comprehensive review. *Annals of Medicine*, 55(2), Article 2295398. <https://doi.org/10.1080/07853890.2023.2295398>
- Ram, R. (2025). Protecting New Zealand’s agricultural legacy through education. *Waikato Journal of Education*, SE-Articles. <https://doi.org/10.15663/wje.a971>
- Rebollada-Merino, A., et al. (2024). Detection of Brucella in Dermacentor ticks of wild boar with brucellosis. *Transboundary and Emerging Diseases*, 2024, Article 6618287. <https://doi.org/10.1155/2024/6618287>
- Riabi, H. R. A., et al. (2017). Epidemiological features of human brucellosis prevalence in southern cities of Khorasan Razavi, Iran. *Zahedan Journal of Research in Medical Sciences*, 19(4), e7911. <https://doi.org/10.5812/zjrms.7911>
- Rossetti, C. A., et al. (2022). Comparative review of brucellosis in small domestic ruminants. *Frontiers in Veterinary Science*, 9, Article 887671. <https://doi.org/10.3389/FVETS.2022.887671>
- Salem, L. M. A., et al. (2014). Serological diagnosis of brucellosis by using simple and rapid field tests with emphasis on some possible risk factors in humans. *Global Veterinaria*, 12(3), 320–325. <https://doi.org/10.5829/idosi.gv.2014.12.03.82146>
- Sharma, H. K., et al. (2016). Seroprevalence of human brucellosis in and around Jammu, India, using different serological tests. *Veterinary World*, 9(7), 742–746.
- Shen, G., et al. (2025). The Brucella strain caused an infant’s Brucella meningitis. *Biosafety and Health*, 7(2), 117–121. <https://doi.org/10.1016/j.bsheal.2025.03.002>
- Shome, R., et al. (2017). Prevalence and risk factors for brucellosis among veterinary healthcare professionals. *Pathogens and Global Health*, 111(5), 234–239. <https://doi.org/10.1080/20477724.2017.1345366>
- Tumwine, G., et al. (2015). Human brucellosis: Sero-prevalence and associated risk factors in agro-pastoral communities of Kiboga District, Central Uganda. *BMC Public Health*, 15, Article 900. <https://doi.org/10.1186/s12889-015-2242-z>
- Wang, L., et al. (2025). Mapping the research landscape of immune response in human brucellosis: A bibliometric analysis. *Frontiers in Microbiology*, 16, Article 1583520. <https://doi.org/10.3389/fmicb.2025.1583520>
- World Health Organization. (2020, July 29). *Brucellosis*. <https://www.who.int/news-room/fact-sheets/detail/brucellosis>
- Zeb, S., et al. (2025). Antimicrobial-resistant Brucella spp. prevail in raw milk and animal feces of different livestock farms. *BMC Microbiology*, 25, Article 231. <https://doi.org/10.1186/s12866-025-03930-8>